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## ARTIGO DE REVISÃO

### ***Helicobacter pylori pathogenicity genes, cytokine polymorphisms and environmental factors affect the development of gastric diseases: an overview***

***Genes de patogenicidade de Helicobacter pylori, polimorfismos de citocinas e fatores ambientais afetam o desenvolvimento de doenças gástricas: uma visão geral***

Júlia Silveira Vianna,<sup>1</sup> Pedro Eduardo Almeida da Silva,<sup>1,2</sup> Ivy Bastos Ramis<sup>2</sup>

<sup>1</sup>Universidade Federal de Pelotas (UFPel), Pelotas, RS, Brasil.

<sup>2</sup>Universidade Federal do Rio Grande (FURG), Rio Grande, RS, Brasil.

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jusvianna@hotmail.com

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*Bacterial genes;*

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*Diseases;*

*Epidemiology.*

## RESUMO

**Justificativa e Objetivos:** *Helicobacter pylori* é uma bactéria Gram negativa que coloniza o estômago de aproximadamente 50% da população humana mundial. Este microrganismo é o principal agente causal de gastrite e um importante fator de risco para o desenvolvimento de úlcera péptica e carcinoma gástrico. Os fatores que determinam essa diversidade de manifestações clínicas permanecem incertos, mas podem estar relacionados com a interação dos fatores bacterianos, sistema imune do hospedeiro e variáveis ambientais. O objetivo desta revisão é fornecer uma visão geral destes fatores que influenciam na susceptibilidade a desordens severas de infecção por *H. pylori*. **Metodo:** Para isso, foram selecionados artigos originais e de revisão através da pesquisa nas bases de dados bibliográficos PubMed, Portal de Periódicos CAPES e SCIELO. **Resultados:** *H. pylori* possui um conjunto de fatores de patogenicidade, tais como *cagA*, *vacA*, *iceA*, *babA*, para colonizar a mucosa gástrica e estabelecer infecção crônica. Estes fatores bacterianos são agentes essenciais em modular a resposta imune envolvida na iniciação da carcinogênese gástrica. Os fatores genéticos do hospedeiro contribuem para regular a resposta inflamatória e para o agravamento da lesão da mucosa gástrica uma vez que a infecção gástrica por *H. pylori* induz a produção de várias citocinas pró e anti-inflamatórias no hospedeiro. O papel prejudicial dos fatores ambientais está relacionado com as precárias condições socioeconômicas, com o consumo de sal, com o tabagismo e com o consumo de álcool. **Conclusão:** Ao decifrar as regras deterministas - se houver - dessa interação entre fatores da bactéria, do hospedeiro e variáveis ambientais, será possível prever, tratar e, finalmente, prevenir graves doenças gastroduodenais.

## ABSTRACT

**Background and Objectives:** *Helicobacter pylori* is a Gram-negative bacterium that colonizes the stomach of approximately 50% of the world's human population. This microorganism is the major causal agent of gastritis and is an important risk factor for the development of peptic ulcer disease and gastric carcinoma. The factors that determine these diverse clinical outcomes are subject to continuous investigations and is thought to be determined by interaction of bacterial factors, host immune system and environmental variables. The aim of this review is to provide an overview of these factors that influence susceptibility to severe outcomes of *H. pylori* infection. **Methods:** For this, original and review articles were selected by searching the PubMed, CAPES Portal Journals and SCIELO bibliographic databases. **Results:** *H. pylori* possesses a set of pathogenicity factors, such as *cagA*, *vacA*, *iceA*, *babA* for colonizing the gastric mucosa and establishing a chronic infection. These bacterial factors are essential players in modulating the immune response involved in the initiation of the carcinogenesis in the stomach. Host genetic factors contribute to the regulation of the inflammatory response and in the

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aggravation of mucosal damage once the gastric infection with *H. pylori* induces the mucosal production of various pro- and anti-inflammatory cytokines in the host. The harmful role of environmental factors is related to poor socioeconomic conditions, salt intake, smoking and alcohol consumption. **Conclusion:** By deciphering the deterministic rules – if any – of this interplay between factors of the bacterium, host and environmental variables, we will eventually be able to predict, treat, and ultimately prevent serious gastroduodenal diseases.

## INTRODUCTION

*Helicobacter pylori* is a micro-aerophilic spiral-shaped flagellar Gram-negative bacterium that colonizes the gastric mucosa of approximately 50% of the world's human population.<sup>1</sup> The prevalence in developing countries is estimated between 60–90%, and in the developed world is between 25–35%.<sup>2</sup>

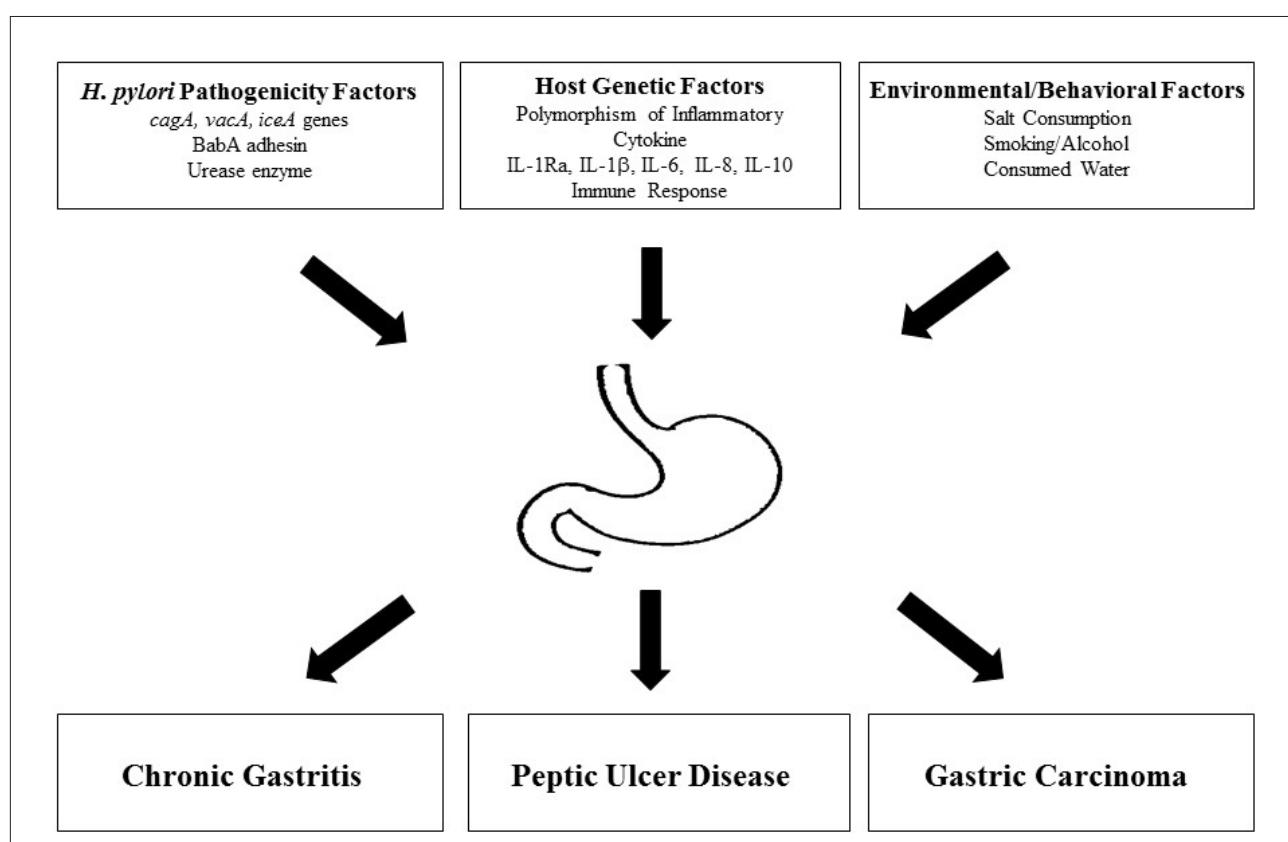
*H. pylori* induces a chronic gastric inflammation, which is asymptomatic in the majority of the patients. However, it is estimated that 15 to 20% of *H. pylori* infected individuals will develop peptic ulcer, and approximately 1% will develop gastric cancer.<sup>3</sup> The interaction of bacterial pathogenicity factors, host factors and environmental and lifestyle factors, determine the severity of gastric damage and the clinical outcome of *H. pylori* infection (Figure 1).<sup>4</sup>

The *H. pylori* genome has a high plasticity, therefore, genomic variations of the strains may be responsible for the coding of different pathogenic factors capable of determining various types of gastric lesions in the host.<sup>5</sup>

It has been described as biomarkers of pathogenicity of *H. pylori*, *cagA* (cytotoxin-associated gene A), *vacA* (vacuolating cytotoxin), *iceA* (induced by contact with epithelium), *babA* (blood-group antigen-binding adhesin A), among others genes.<sup>6-9</sup>

The host's immune system plays an important role in the pathogenesis of gastroduodenal disorders by regulating the nature and the intensity of the inflammatory response to infection by *H. pylori*. Factors such as cytokines are involved in the initiation and regulation of this response.<sup>10</sup> The inflammatory cells that are recruited to the gastric mucosa during infection produce several pro- and anti-inflammatory cytokines.<sup>11</sup> In this sense, it is noteworthy that the polymorphism in the genes encoding interleukins (*IL*) 1 $\beta$ , 1RN, 6, 8 and 10 are associated with increased susceptibility to gastric diseases.<sup>12-17</sup>

The combined presence of environmental factors, such as, household crowding, inadequate eating habits, low socioeconomic conditions, poor hygienic, excessive alcohol consumption and smoking are related with *H. pylori* infection, especially when one takes into account



**Figure 1.** A schematic to explain why only certain individuals infected by *H. pylori* develop more severe gastric diseases. The interplay between bacterial, host and environmental/behavioral factors determine the different clinical outcomes of *H. pylori*-infected patients.

their main routes of transmission, oral-oral and fecal-oral. Besides, factors as smoking, eating habits and alcohol consumption have been associated with a higher risk of the development of peptic ulcer and gastric cancer in people infected by *H. pylori*.<sup>18-20</sup>

The aim of this review is to provide an overview of bacterial, environmental, and host factors that influence susceptibility to severe outcomes of *H. pylori* infection.

## METHODS

Original and review articles were selected by searching the PubMed (Literature of International Health Sciences), CAPES Portal Journals and SCIELO bibliographic databases. The selection of the articles was based on two criteria: 1) published between the years 1993 and 2015; 2) articles whose authors contributed with important insights into the issues, regardless of the year of publication. The following descriptors were used: *Helicobacter pylori*, pathogenicity genes, host genetic polymorphism, cytokine polymorphism, environmental variables, gastroduodenal diseases and epidemiology.

## RESULTS AND DISCUSSION

### ***Helicobacter pylori* infection: From gastric inflammation to gastric carcinogenesis**

The clinical manifestations of *H. pylori* infection can be chronic gastritis; peptic ulcer; adenocarcinoma; and gastric lymphoma mucosa-associated lymphoid tissue (MALT).<sup>21,22</sup> Firstly, the infection of the stomach by *H. pylori* induces inflammation of the gastric mucosa (gastritis) and is estimated that occur in the most patients infected by *H. pylori*.<sup>23</sup> *H. pylori* infection may be associated with decreased or increased gastric acid secretion, depending on the distribution and severity of gastritis.<sup>4</sup>

The interplay of gastritis and acid secretion are key determinants in disease outcomes such as peptic ulcer and/or gastric cancer.<sup>23</sup> Ulceration corresponds to the loss of mucosal integrity as the result of disequilibrium between defensive mucosa-protective factors (mucus bicarbonate layer and phospholipid surfactant layer) and aggressive injurious factors (strong acid and high proteolytic activity in gastric secretions). When ulcers develop in the acid-peptic environment of the gastroduodenum, they are called peptic ulcer disease.<sup>24</sup> The development of duodenal ulcers is associated with gastritis and with increased gastric acid production, whereas the development of gastric ulcers is related with gastritis and with gastric reduced acid production, conditions that predispose also the developing of gastric atrophy (loss of gastric glands), intestinal metaplasia (the replacement of glandular and/or foveolar epithelium by intestinal epithelium), and finally gastric cancer.<sup>25,26</sup>

In relation to gastric cancer, although its etiology is multifactorial, *H. pylori* is the main cause of gastric carcinogenesis and was categorized in 1994 as a carcinogen I

by the International Cancer Research Agency (IARC).<sup>27</sup> *H. pylori* infection promotes gastric carcinogenesis by at least five different mechanisms: (1) increased endogenous DNA damage and decrease repair activities, (2) induction of mutations in the mitochondrial DNA, (3) generation of a phenotype that induces mutations in the nuclear genome (4) disruption of the balance between cell proliferation and apoptosis and (5) induction of an intense gastric inflammatory response that lasts over decades that produces chronic oxidative stress and adaptive changes in gastric epithelial and immune cell pathobiology.<sup>28</sup>

The gastric cancer classification by system Lauren-classifies gastric adenocarcinoma in intestinal or diffuse.<sup>29</sup> The intestinal-type is more frequently observed in older patients and it is linked closely to environmental and dietary risk factors. The pathogenesis of the intestinal type includes a sequence of events that begins with *H. pylori*-induced superficial gastritis, progressing towards chronic atrophic gastritis, intestinal metaplasia, dysplasia and finally gastric cancer. In diffuse type the patients typically are younger associated with host genetic factors and is generally less common, with a worse prognosis than those with the intestinal type. The pathogenesis of diffuse type is poorly understood, although *H. pylori* infection is also a predisposing factor.<sup>30</sup> Anatomically, gastric adenocarcinomas are classified as proximal (cardia) and distal (noncardia). The longstanding inflammatory response against *H. pylori* in the gastric mucosa may cause sustained tissue injury leading to the development of distal gastric adenocarcinoma.<sup>31</sup>

Lastly, there is a pathogenic link between *H. pylori* infection and MALT lymphoma, so much that, according to the Maastricht III Consensus Report, *H. pylori* eradication is the treatment of first choice for *H. pylori* infected individuals with stage I low grade gastric MALT lymphoma.<sup>32</sup>

### **Role of bacterial pathogenicity genes in the gastric disorders**

Determination of pathogenicity genes may provide information regarding to the prognosis of infection by *H. pylori*.<sup>33</sup> The first gene of pathogenicity identified in *H. pylori* was *cagA*. This gene is highly prevalent in strains from different geographic areas.<sup>6,34</sup> The *cagA* gene is located at one end of a 40-kb DNA insertion called *cag* pathogenicity island (*cagPAI*). This gene is secreted through the type IV secretion system (T4SS) and then transported to host cells.<sup>35</sup> Once inside gastric epithelial cells, the CagA undergoes tyrosine phosphorylation in its repeat region of five amino acids Glu-Pro-Ile-Tyr-Ala (EPIYA). This phosphorylation is mediated by Src kinase, a cytoplasmic protein tyrosine kinase. CagA tyrosine phosphorylated subsequently binds to Src homology 2 (SH2) domain, that contain host cell proteins, such as, the tyrosine phosphatase SHP-2, the C-terminal Src kinase (CSK) and the adapter protein Crk. This results in cytoskeletal reorganization and cell elongation-a phenotype that leads to the dispersion of cells and morphological changes for "hummingbird phenotype".<sup>36</sup> The *cagA*-SHP-2 interaction constitutes the biological basis of *cagA* as

a pathogenicity factor, due to induction of abnormal proliferation and movement of gastric epithelial cells, changes that may eventually lead to gastric atrophy and gastric cancer.<sup>37</sup>

The CagA protein of *H. pylori* acts as a highly immunogenic antigen. The structure of the *cagA* gene reveals a 5' highly conserved region and a 3' region containing a variable number of repeat sequences, which leads to variation in the length of the protein and can result in diverse host responses, including different degrees of inflammatory response.<sup>5</sup> These variations in the repeat region of *cagA*EPIYA have also been associated with pathogenicity of *H. pylori*. Four distinct types of EPIYA have been identified and classified as, EPIYA-A, -B, -C and -D. CSK specifically binds to the tyrosine-phosphorylated EPIYA-A or -B, whereas SHP-2 specifically binds to the tyrosine-phosphorylated EPIYA-C or -D.<sup>36</sup> *cagA* strains of *H. pylori* found in the western world typically contain EPIYA-A, -B and -C in the EPIYA repetition region in the C-terminal. In contrast, the *cagA* strains of the East Asian have EPIYA-A, -B and a specific sequence Asian CagAEPIYA-D.<sup>38</sup> The number of segments EPIYA-C in *H. pylori* strains from western countries have shown to influence the degree of pathogenicity as well as the oncogenic potential, and may serve as a marker for identifying high-risk populations.<sup>39</sup> This occurs because the presence of EPIYA-C segments of the CagA protein appears to significantly contribute to the transcriptional activation of IL-8, through activation of NF- $\kappa$ B (factor nuclear kappa B).<sup>40</sup> IL-8 plays a crucial role by chemo attracting and activating neutrophils to the site of infected gastric mucosa.<sup>41</sup> Batista et al. showed that increasing of the number of segments EPIYA-C was associated with precancerous gastric lesions and with decreased serum levels of pepsinogen I, which reflects the functional and morphological status of the gastric mucosa.<sup>42</sup> Also, it was observed that infection by *cagA*-positive *H. pylori* strains harboring multiple EPIYA-C repeats is associated with the presence of peptic ulcer.<sup>43</sup>

Another important disease-associated pathogenicity genes of *H. pylori* are *vacA*, *iceA* and *babA*.<sup>7-9</sup> The *vacA* gene encodes a vacuolating cytotoxin (VacA), which can damage the gastric epithelial cells due to induce the formation of cytoplasmic vacuoles.<sup>44</sup> The gene consists of three variable regions: the signal region (*s*- encoding the signal peptide), with two alleles, *s1* (subtypes *s1a*, *s1b*, *s1c*) and *s2*; the medium region (*m*), with the alleles *m1* and *m2*; and the intermediate region (*i*), containing the alleles *i1* and *i2*.<sup>44,45</sup> The combination of alleles of the *s*, *m* and *i* regions determines the vacuolating cytotoxin production and is associated with the pathogenicity of the bacterium. In general, the strains containing *s1/m1/i1* alleles produce large amounts of vacuolating cytotoxin, reason where by this genotype seems to be associated with more severe pathologies, as peptic ulcer and gastric cancer. In contrast, strains of type *s1/m2* and *s2/m2/i2* have moderate and none cytotoxic activity, respectively.<sup>7,46</sup> A recent study showed that the best markers of gastric cancer and duodenal ulcer were the *vacA s1* and *i1* genotypes, and that the *s* and *i* regions were the key determinants of vacuolating cytotoxin activity.<sup>47</sup>

Another study found that strains which carried the *vacA s1/m1* gene showed significant associations with severe chronic gastritis.<sup>48</sup> The *iceA* gene has two alleles: *iceA1* and *iceA2*. The expression of *iceA1* is regulated by the contact of *H. pylori* with gastric epithelial cells and is associated with peptic ulcer and gastric cancer, whereas expression of *iceA2* is related to asymptomatic gastritis.<sup>8</sup> A study in southern Brazil showed that the *iceA1* allele was related to erosive gastritis, whereas the *iceA2* allele was associated with enanthematos gastritis.<sup>49</sup> The *babA* gene encodes a membrane protein, named BabA adhesin, which binds to the Lewis b blood group antigens in gastric cells. The *babA* gene is a factor of adhesion, one of the characteristics of *H. pylori* that assists in establishing persistent colonization of the gastric epithelium and contributes to its pathogenicity, by to allow intimate contact between bacterium and epithelium, and facilitate the release of factors pathogenicity. Although three alleles of the *babA* gene have been identified (*babA1*, *babA2* and *babB*), only the *babA2* gene product is able to bind to Lewis b antigen.<sup>50</sup> A recent study showed that the *babA2* gene correlated positively with bacterial density score, activity of inflammation and chronic inflammation of gastric mucosa.<sup>51</sup>

### Influence of cytokines genes polymorphisms in the gastric diseases

*H. pylori* induces gastric lesions that start by chronic inflammation in the gastric mucosa that is mediated by an array of pro- and anti-inflammatory cytokines.<sup>52</sup> Genetic polymorphisms in genes that codify proinflammatory cytokines such as *IL-1 $\beta$* , *IL-6* and *IL-8*; and anti-inflammatory cytokines like *IL-1RN* and *IL-10*, directly influence inter-individual variation in the magnitude of the cytokine response, and this clearly contributes to an individual's ultimate clinical outcome.<sup>53</sup> In general, in the *H. pylori* infection, the gastric inflammation is increased and the acid inhibition is potent in patients with high producer alleles of pro-inflammatory cytokines and low producer alleles of anti-inflammatory cytokines, which results in a higher risk for the development of atrophic gastritis, gastric ulcer or gastric cancer. On the other hand, low producer allele carriers of pro-inflammatory cytokines and high producer allele carriers of anti-inflammatory cytokines have decreased inflammation and weak acid inhibition, resulting in a mild gastric inflammation.<sup>54</sup>

The family of the *IL-1* gene contains the genes *IL-1A*, *IL-1B* and *IL-1RN*.<sup>55</sup> *IL-1B* encodes *IL-1 $\beta$* , a pro-inflammatory cytokine and a powerful inhibitor of gastric acid secretion, which plays an important role for initiating and amplifying the inflammatory response to *H. pylori* infection.<sup>56</sup> There are three single nucleotide polymorphisms (SNPs) in the *IL-1 $\beta$*  gene: a T-C base transition at position -31 and C-T base transitions at positions -511 and +3954 from the transcriptional start site.<sup>57</sup> The presence of the T allele at positions -511 and +3954; and of C allele at position -31 is associated with high levels

of *IL-1* secretion.<sup>25</sup> It has been shown that *IL-1 $\beta$ -31*, *IL-1 $\beta$ -511* and *IL-1 $\beta$ +3954* polymorphisms are related with hypochlorhydria, chronic atrophic gastritis, gastric ulcer and gastric cancer in response to *H. pylori* infection.<sup>52,58</sup> The *IL-1RN* encodes the IL-1 receptor antagonist (IL-1ra), an anti-inflammatory cytokine that competitively binds to IL-1 receptors and thereby modulates the potentially damaging effects of IL-1.<sup>12</sup> *IL-1RN* play a decisive role in modulating the risk of developing hypochlorhydria, gastric atrophy and gastric cancer in the presence of *H. pylori* infection.<sup>59</sup> *IL-1RN* gene contains an 86-bp variable number of tandem repeats polymorphism (VNTR) in intron 2, which leads to the presence of 5 different alleles: allele 1 (4 repeats), allele 2 (2 repeats), allele 3 (5 repeats), allele 4 (3 repeats), and allele 5 (6 repeats).<sup>60</sup> The 4-repeats (*IL-1RN\*1*) and 2-repeats (*IL-1RN\*2*) alleles are the most common, whereas the others account for less than 5%.<sup>61</sup> The allele 2 (*IL-1RN\*2*) is associated with enhanced *IL-1 $\beta$*  production.<sup>62</sup> Studies in different populations revealed association between individual's carriers of the allele 2 and increased risk of developing chronic gastritis, gastric ulcer and gastric cancer.<sup>63,64</sup>

*IL-6* is a pro-inflammatory cytokine that acts as an inflammatory mediator and endocrine regulator, playing an important role in host defense mechanisms as a messenger between innate and adaptive systems.<sup>65</sup> Moreover, it is involved in the regulation of various cellular functions, such as, proliferation, apoptosis and angiogenesis.<sup>66</sup> A nucleotide change from G to C at position -174 in the promoter region of the *IL-6* gene was described. This SNP may result in inter-individual variation in transcription and expression of the *IL-6*; and therefore influence an individual's susceptibility to a diverse range of diseases. Mucosal *IL-6* levels are elevated in *H. pylori*-associated gastritis and diminished after eradication of the infection.<sup>55</sup> In 2012, Liu et al. showed an increased cancer risk for individuals with the CC genotype compared to those carrying the GG genotype in African populations.<sup>67</sup>

The pro-inflammatory factor *IL-8* is recognized as a neutrophil and lymphocyte chemotactic factor and inductor of cell proliferation, migration and angiogenesis.<sup>54,68</sup> Gastric mucosal levels of *IL-8* increase significantly after *H. pylori* infection and parallel to the severity of gastritis.<sup>69</sup> The *IL-8* gene contains a SNP (T-A base transition) at position -251, in the proximal promoter region. The *IL-8-251A* allele affects *IL-8* gene transcription and tended to be associated with increased *IL-8* production by gastric cells and with more severe inflammation.<sup>53</sup> In recent years, a number of studies examining the *IL-8-251T/A* SNP showed varying results. Recent meta-analyses suggest that *IL-8-251T/A* SNP is associated with increased peptic ulcer and gastric cancer risks among Asians.<sup>70</sup> However, a study examining European patients did not find the correlation between the *IL-8-251T/A* SNP and gastric cancer.<sup>71</sup> Moreover, a Brazilian study found that individuals with A/A genotype may have protective effect for gastric cancer.<sup>72</sup> These contrasting results suggest that *IL-8-251T/A* SNP may be associated differently with gastric diseases depending on the ethnicity.

*IL-10* is a pleiotropic anti-inflammatory cytokine produced by activated immune cells.<sup>73</sup> It is capable of inhibiting the production of pro-inflammatory cytokines like IFN- $\gamma$ , IL-1, IL-2, IL-3, IL-6, IL-8, TNF- $\alpha$ , and GM-CSF and inducing B-cell proliferation and differentiation.<sup>74,75</sup> A C-T base transition located at positions -819, and C-A base transition at position -592 have been identified in the 5' flanking region of the *IL-10* gene. These SNPs are related to different serum level of *IL-10* in vivo.<sup>75,76</sup> The C allele at positions -819 and -592 has been associated with high *IL-10* production and the T allele at position -819 and the A allele at position -592 has been related with low production.<sup>77</sup> The low *IL-10* production in patients infected with *H. pylori* results in increased gastric inflammation intensity, hypochlorhydria and increased risk of gastric atrophy and gastric cancer.<sup>74</sup>

### Relationship of environmental factors with the *H. pylori* infection

The knowledge of the epidemiology and mode of transmission of *H. pylori* is important to prevent its spread, since this bacterium is present in nearly half the world's population. Such knowledge may be useful in identifying high-risk populations, especially in areas with higher rates of peptic ulcer and gastric cancer.<sup>5,78,79</sup>

The transmission of *H. pylori* occurs from person to person by oral-oral (through saliva, dental plaque, backflow of gastric contents) or fecal-oral (through water contaminated) routes.<sup>78</sup> Yu et al. (2015) showed that dental plaque is risk factor of oral *H. pylori* infection.<sup>80</sup> In Brazil, studies showed that mothers infected with *H. pylori* represent a greater risk factor to infected their children; and that breastfeeding does not protect against acquisition of *H. pylori*, conversely, an infected mother can transmit of the disease to the child.<sup>81,82</sup>

The theory that the major risk factors for infection are poor socioeconomic and hygienic conditions beginning in childhood was supported by Chen et al. (2014).<sup>83</sup> Childhood infections, such as, tonsillitis, diarrhea and diphtheria, are associated with decreased secretion of gastric acid, as well as with malnutrition. Regions where childhood infections and malnutrition are common provide the ideal environment for *H. pylori* colonization.<sup>84</sup>

There is a strong correlation between poor socio-economic conditions and *H. pylori* infection. In general, inadequate sanitation practices, low family income, poor hygiene and household crowding may be related to a higher prevalence of this infection.<sup>19</sup> The transmission of *H. pylori* occurs from infected mothers to their offspring and among siblings.<sup>85</sup> A study, with Brazilian children, showed that the number of siblings and nursery attendance were positively associated with *H. pylori* infection and also that the house location (served by paved road or not) is a potential risk factor for infection, indicating that contaminated soil may be a source of infection, and concluding that the *H. pylori* infection in children is highly related to poor hygiene and crowded conditions.<sup>86</sup> A study conducted in six Latin-American countries also reported that crowding was positively associated with *H. pylori*.

*pylori* infection and suggested the occurrence of repeated transmission of *H. pylori* (same or different strain) between the individuals who live in the same household, due to a greater opportunity of personal contact.<sup>19</sup>

In addition to the factors described above, other variables, such as smoking, alcohol consumption and diet may influence the infection by *H. pylori*. The smoking can be involved in the transmission of infection due the handling and sharing of cigarettes among smokers.<sup>87</sup> A population-based prospective study with Japanese men suggested that cigarette smoking and *H. pylori* infection are significant risk factors for gastric cancer.<sup>88</sup> The strong oxidising effect of cigarette smoke is able to dramatically induce oxidative DNA damage.<sup>4</sup>

Regarding alcohol consumption, Sánchez-cuen et al (2014) found that the risk of infection by *H. pylori* is 1.45 times higher in people who consume alcohol who do not consume it.<sup>18</sup> It is postulated that alcohol consumption facilitates *H. pylori* infection presumably by damaging the gastric mucosa and/or promoting *H. pylori* adherence to gastric mucosa.<sup>68</sup>

Another factor that influences the infection by *H. pylori* is the diet that may provide noxious agents that contribute to *H. pylori* pathogenicity. The high salt concentration in the stomach destroys the mucosal barrier, favors colonization by *H. pylori*, and leads to inflammation and damage-causing gastritis and diffuse erosion.<sup>4</sup> Inadequate dietary habits are associated with intense neutrophilia, higher degree of inflammation and with the development of gastric cancer.<sup>20</sup>

Lastly, a recent study demonstrated a statistically significant increase in *H. pylori* infection among betel chewers compared to those who did not chew betel. This observation supports the hypothesis that betel chewing may affect periodontal health and thus predispose individuals to colonization with *H. pylori*.<sup>83</sup>

## CONCLUSION

There are evidences that genes of bacterial pathogenicity, such as, *cagA*, *vacA s1/m1/i1*, *iceA1*, *babA2*; host genetic polymorphisms, such as, *IL-1RN* \*2, *IL-1β* (-31C) (-511T) (+3954T), *IL-8* (-251A), *IL-10* (-592A) (-819T); and environmental factors as smoke, alcohol consumption and diet play a major role in inducing severe gastric disorders. Future research must focus on developing a diagnostic tool for detect *H. pylori* and also bacterial and host genes associated with a higher severity of gastric disease. The identification of patients infected with *H. pylori* that present a high risk of develop peptic ulcer or gastric cancer may to determine the most appropriate therapeutic intervention and the better treatment.

## REFERENCES

1. Wen S, Moss SF. Helicobacter pylori virulence factors in gastric carcinogenesis. *Cancer Lett* 2009;282(1):1-8. doi: 10.1016/j.canlet.2008.11.016.
2. de Vries AC, Kuipers EJ. Helicobacter pylori infection and nonmalignant diseases. *Helicobacter* 2010;15(1):29-33. doi: 10.1111/j.1523-5378.2010.00775
3. Menon R, Riera A, Ahmad A. A global perspective on gastrointestinal diseases. *Gastroenterol. Clin North Am* 2011; 40(2):427-439. doi: 10.1016/j.gtc.2011.03.002
4. Izzotti A, Durando P, Ansaldi F, et al. Interaction between Helicobacter pylori, diet, and genetic polymorphisms as related to non-cancer diseases. *Mutat Res* 2009;667(1-2):142-157. doi: 10.1016/j.mrfmmm.2009.02.002
5. Ladeira MSP, Salvadori DMF, Rodrigues MAM. Biopathology of Helicobacter pylori. *J Bras Patol Med Lab* 2003;39(1):335-342. doi: 10.1590/S1676-24442003000400011
6. Covacci A, Censini S, Bugnoli M, et al. Molecular characterization of the 128-kDa immunodominant antigen of Helicobacter pylori associated with cytotoxicity and duodenal ulcer. *Proc Natl Acad Sci USA* 1993;90(12):5791-5795.
7. Atherton JC, Cao P, Peek Jr. RM, et al. Mosaicism in vacuolating cytotoxin alleles of Helicobacter pylori: association of specific *vacA* types with cytotoxin production and peptic ulceration. *J Biol Chem* 1995;270(30):17771-17777. doi: 10.1074/jbc.270.30.17771
8. Peek Jr RM, Thompson SA, Donahue JP, et al. Adherence to gastric epithelial cells induces expression of a Helicobacter pylori gene, *iceA*, that is associated with clinical outcome. *Proc Assoc Am Physicians* 1998;110(6):531-544.
9. Gerhard M, Lehn N, Neumayer N, et al. Clinical relevance of the Helicobacter pylori gene for blood-group antigen-binding adhesin. *Proc Natl Acad Sci USA* 1999;96(22):12778-12783. doi: 10.1073/pnas.96.22.12778
10. Li C, Xia HH, Xie W, et al. Association between interleukin-1 gene polymorphisms and Helicobacter pylori infection in gastric carcinogenesis in a Chinese population. *J Gastroenterol Hepatol* 2007;22(2):234-239. doi: 10.1111/j.1440-1746.2006.04379
11. Horvath Jr DJ, Radin JD, Cho SH, et al. The Interleukin-17 Receptor B Subunit Is Essential for the Th2 Response to Helicobacter pylori, but Not for Control of Bacterial Burden. *PLoS ONE* 2013; 8(3):1-8. doi: 10.1371/journal.pone.0060363
12. Arend WP, Malyak M, Guthridge CJ, et al. Interleukin-1 receptor antagonist: role in biology. *Annu Rev Immunol* 1998;16(1):27-55. doi: 10.1146/annurev.immunol.16.1.27
13. Bodger K, Wyatt JI, Heatley RV. Gastric mucosal secretion of interleukin-10: relations to histopathology, Helicobacter pylori status, and tumour necrosis factor-alpha secretion. *Gut* 1997; 40(6): 739-744. doi: 10.1136/gut.40.6.739
14. Crabtree JE, Lindley IJ. Mucosal interleukin-8 and Helicobacter pylori-associated gastroduodenal disease. *Eur J Gastroenterol Hepatol* 1994; 6(1): S33-S38.
15. El-Omar EM, Carrington M, Chow WH, et al. Interleukin-1 polymorphisms associated with increased risk of gastric cancer. *Nature* 2000;404(6776):398-402. doi: 10.1038/35006081
16. Gatti LL, Tunes MZ, Lábio RW, et al. Interleukin-6 polymorphism and Helicobacter pylori infection in Brazilian adult patients with chronic gastritis. *Clin Exp Med* 2005;5(3):112-116. doi: 10.1007/s10238-005-0074-3
17. Kang JM, Kim N, Lee DH, et al. The effects of genetic

- polymorphisms of IL-6, IL-8, and IL-10 on Helicobacter pylori-induced gastroduodenal diseases in Korea. *J Clin Gastroenterol* 2009;43(5):420-428. doi: 10.1097/MCG.0b013e318178d1d3
18. Sánchez-Cuén JA, Cabrales ABI, Magaña GB, et al. Infección por Helicobacter pylori y su asociación con el consumo de alcohol. Estudio de casos y controles. *Rev Gastro de México* 2013;78(3): 144-150. doi: 10.1016/j.rgmx.2013.06.003
19. Porras C, Nodora J, Sexton R, et al. Epidemiology of Helicobacter pylori infection in six Latin American countries (SWOG Trial S0701). *Cancer Causes Control* 2013;24(2):209-215. doi: 10.1007/s10552-012-0117-5
20. Vinagre RMDF, Vilar-e-Silva A, Fecury AA, et al. Role of Helicobacter pylori infection and lifestyle habits in the development of gastroduodenal diseases in a population from the Brazilian Amazon. *Arq Gastroenterol* 2013;50(3):170-174. doi: 10.1590/S0004-28032013000200030
21. Kandulski M, Selgrad P, Malfertheiner. Helicobacter pylori infection: a clinical overview. *Dig Liver Dis* 2008;40(8):619-626. doi: 10.1016/j.dld.2008.02.026
22. Noto JM, Peek RM. Helicobacter pylori: an overview. *Methods Mol Biol* 2012;921(1):7-10. doi: 10.1007/978-1-62703-005-22
23. Malfertheiner P. The intriguing relationship of Helicobacter pylori infection and acid secretion in peptic ulcer disease and gastric cancer. *Dig Dis* 2011;29(5):459-464. doi: 10.1159/000332213
24. Tytgat GN. Etiopathogenetic principles and peptic ulcer disease classification. *Dig Dis Magdeburg* 2011;29(5):454-458. doi: 10.1111/j.1440-1746.1991.tb01469
25. Zabaleta J. Multifactorial Etiology of Gastric Cancer. *Methods Mol Biol* 2012;863(1):411-435. doi: 10.1007/978-1-61779-612-8\_26
26. Rugge M, Capelle LG, Cappellosso R, et al. Precancerous lesions in the stomach: from biology to clinical patient management. *Best Pract Res Clin Gastroenterol* 2013;27(2):205-223. doi: 10.1016/j.bpg.2012.12.007
27. World Health Organization, International Agency for Research on Cancer. Schistosomes, Liver Flukes and Helicobacter pylori. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans 1994;61:177-240.
28. Costa AC, Figueiredo C, Touati E. Pathogenesis of Helicobacter pylori infection. *Helicobacter* 2009;14(1):15-20. doi: 10.1111/j.1523-5378.2009.00702
29. Lauren P. The two histological main types of gastric carcinoma: diffuse and so-called intestinal-type carcinoma. An attempt at a histo-clinical classification. *Acta Pathol Microbiol Scand* 1965; 64(1): 31-49.
30. Jang BG, Kim WH. Molecular pathology of gastric carcinoma. *Pathobiology* 2011;78(6):302-310. doi: 10.1159/00032170.
31. Piazuelo MB, Epplein M, Correa P. Gastric cancer: an infectious disease. *Infect Dis Clin North Am* 2010;24(4):853-869. doi: 10.1016/j.idc.2010.07.010
32. Malfertheiner P, Megraud F, O'Morain C, et al. Current concepts in the management of Helicobacter pylori infection: the Maastricht III Consensus Report. *Gut* 2007;56(6):772-81. doi: 10.1136/gutjnl-2012-302084
33. Essawi T, Hammoudeh W, Sabri I, et al. Determination of Helicobacter pylori Virulence Genes in Gastric Biopsies by PCR. *Gastroenterol* 2013;2013(1):1-4. doi: 10.1155/2013/606258
34. Arévalo-galvis A, Trespalacios-Rangel AA, Otero W, et al. Prevalence of cagA, vacA, babA2 and iceA genes in *H. pylori* strains isolated from Colombian patients with functional dyspepsia. *Pol J Microbiol* 2012;61(1):33-40.
35. Censini S, Lange C, Xiang Z, et al. cag, a pathogenicity island of *Helicobacter pylori*, encodes type I-specific and disease-associated virulence factors. *Proc Natl Acad Sci USA* 1996;93(25): 14648-14653.
36. Higashi H, Tsutsumi R, Muto S, et al. SHP-2 tyrosine phosphatase as an intracellular target of *Helicobacter pylori* CagA protein. *Science* 2002;295(5555):683-686. doi: 10.1126/science.1067147
37. Ando T, Goto Y, Maeda O, et al. Causal role of *Helicobacter pylori* infection in gastric cancer. *World J Gastroenterol* 2006; 12(2):181-186. doi: 10.3748/wjg.v12.i2.181
38. Backert S, Tegtmeyer N, Selbach M. The versatility of *Helicobacter pylori* cagA effector protein functions: The master key hypothesis. *Helicobacter* 2010;15(3):163-176. doi: 10.1111/j.1523-5378.2010.00759
39. González I, Romero J, Rodríguez B, et al. High prevalence of virulence-associated genotypes in *Helicobacter pylori* clinical isolates in the Region del Maule, Chile. *Scand J Infect Dis* 2011; 43(8):652-655. doi: 10.3109/00365548.2011.572909
40. Papadakos KS, Sougleris IS, Mentis AF, et al. Presence of Terminal EPIYA Phosphorylation Motifs in *Helicobacter pylori* CagA Contributes to IL-8 Secretion, Irrespective of the Number of Repeats. *PLoS ONE* 2013;8(2):1-11. doi: 10.1371/journal.pone.0056291
41. Salih BA, Guner A, Karademir A, et al. Evaluation of the effect of cagPAI genes of *Helicobacter pylori* on AGS epithelial cell morphology and IL-8 secretion. *Antonie Van Leeuwenhoek* 2014;105(1):179-189. doi: 10.1007/s10482-013-0064-5
42. Batista SA, Rocha GA, Rocha AMC, et al. Higher number of *Helicobacter pylori* CagA EPIYA C phosphorylation sites increases the risk of gastric cancer, but not duodenal ulcer. *BMC Microbiology* 2011;11(61):1-7. doi: 10.1186/1471-2180-11-61
43. Vianna JS, Ramis IB, Halicki PCB, et al. Detection of *Helicobacter pylori*CagA EPIYA in gastric biopsy specimens and its relation to gastric diseases. *Diagn Microbiol Infect Dis* 2015;83(2):89-92. doi: 10.1016/j.diagmicrobio.2015.05.017
44. Van Doorn LJ, Figueiredo C, Sanna R, et al. Expanding allelic diversity of *Helicobacter pylori* vacA. *J Clin Microbiol* 1998;36(9):2597-2603.
45. Rhead JL, Letley DP, Mohammadi M, et al. A new *Helicobacter pylori* vacuolating cytotoxin determinant, the intermediate region, is associated with gastric cancer. *Gastroenterology* 2007;133(3):926-936. doi: 10.1053/j.gastro.2007.06.056
46. Ozbey G, Demirel U, Aygun C, et al. Investigation of the association between clinical outcome and the cag pathogenicity-island and other virulence genes of *Helicobacter pylori* isolates from patients with dyspepsia in Eastern Turkey. *Braz J Microbiol* 2013;44(2). doi: 10.1590/S1517-83822013000400034
47. Memon AA, Hussein NR, Miendje Deyi VY, et al. Vacuolating Cytotoxin Genotypes Are Strong Markers of Gastric Cancer and Duodenal Ulcer-Associated *Helicobacter pylori* Strains: a Matched Case-Control Study. *J Clin Microbiol* 2014;52(8):2984-2989. doi: 10.1128/JCM.00551-14

48. Vaziri F, Peerayeh SN, Alebouyeh M, et al. Diversity of Helicobacter pylori genotypes in Iranian patients with different gastroduodenal disorders. *World J Gastroenterol* 2013;19(34):5685-5692. doi: 10.3748/wjg.v19.i34.5685
49. Ramis IB, Fonseca TL, Moraes EP, et al. Molecular Basis of Pathogenicity in Helicobacter pylori Clinical Isolates. *J Clin Microbiol* 2010;48(10):3776-3778. doi: 10.1128/JCM.00472-10
50. Lima VP, Rabenhorst SHB. Genes Associados à Virulência de Helicobacter pylori. *Rev Bras Cancerol* 2009;5(4):389-396.
51. Homan M, Sterbenc A, Kocjan BJ, et al. Prevalence of the Helicobacter pylori babA2 gene and correlation with the degree of gastritis in infected Slovenian children. *Antonie Van Leeuwenhoek* 2014;106(4):637-45. doi: 10.1007/s10482-014-0234-0
52. Wang SY, Shen XY, Wu CY, et al. Analysis of whole genomic expression profiles of Helicobacter pylori related chronic atrophic gastritis with IL-1B-31CC/-51TT genotypes. *J Dig Dis* 2009;10(2):99-106. doi: 10.1111/j.1751-2980.2009.00367
53. Xue H, Wang YC, Lin B, et al. A Meta-Analysis of Interleukin-10 -592 Promoter Polymorphism Associated with Gastric Cancer Risk. *PLoS ONE* 2012;7(7):1-11. doi: 10.1186/1471-2407-12-102
54. Sugimoto M, Yamaoka Y, Furuta T. Influence of interleukin polymorphisms on development of gastric cancer and peptic ulcer. *World J Gastroenterol* 2010;16(10):1188-1200. doi: 10.3748/wjg.v16.i10.1188
55. Dinarello CA. Biologic basis for interleukin-1 in disease. *Blood* 1996;87(6):2095-2147.
56. Santos JC, Ladeira MSP, Pedrazzoli Jr J, et al. Relationship of IL-1 and TNF- $\alpha$  polymorphisms with Helicobacter pylori in gastric diseases in a Brazilian population. *Braz J Med Biol Res* 2012;45(9):811-817. doi: 10.1590/S0100-879X2012007500099
57. Persson C, Engstrand L, Nyrén O, et al. Interleukin 1-beta gene polymorphisms and risk of gastric cancer in Sweden. *Scand J Gastroenterol* 2009;44(3):339-345. doi: 10.1080/00365520802556015
58. Farshad S, Rasouli M, Jamshidzadeh A, et al. IL-1 $\beta$  (+3953 C/T) and IL-8 (-251 A/T) gene polymorphisms in *H. pylori* mediated gastric disorders. *Iran J Immunol* 2010;7(2): 6-108. <http://dx.doi.org/IJIV7i2A5>
59. Erzin Y, Koksal V, Altun S, et al. Role of hostinterleukin 1betagene (IL-1B) and interleukin 1 receptor antagonist gene (IL-1RN) polymorphisms in clinical outcomes in Helicobacter pylori-positive Turkish patients with dyspepsia. *J Gastroenterol* 2008;43(9):705-710. doi: 10.1007/s00535-008-2220-7
60. Tarlow JK, Blakemore AI, Lennard A, et al. Polymorphism in human IL-1 receptor antagonist gene intron 2 is caused by variable numbers of an 86-bp tandem repeat. *Hum Genet* 1993;91(4):403-404.
61. Kim JJ, Kim N, Hwang S, et al. Relationship of interleukin-1 $\beta$  levels and gastroesophageal reflux disease in Korea. *J Gastroenterol Hepatol* 2013;28(1):90-98. doi: 10.1111/j.1440-1746.2012.07274
62. Santtila S, Savinainen K, Hurme M. Presence of the IL-1RA allele 2 (IL1RN\*2) is associated with enhanced IL-1beta production in vitro. *Scand J Immunol* 1998; 47(3):195-198. doi: 10.1046/j.1365-3083.1998.00300
63. Persson C, Canedo P, Machado JC, et al. Polymorphisms in inflammatory response genes and their association with gastric cancer: A HuGE systematic review and meta-analyses. *Am J Epidemiol* 2011;173(3):259-270. doi: 10.1093/aje/kwq370
64. De Oliveira JG, Rossi AF, Nizato DM, et al. Profiles of gene polymorphisms in cytokines and Toll-like receptors with higher risk for gastric cancer. *Dig Dis Sci* 2013;58(4):978-988. doi: 10.1007/s10620-012-2460-5
65. Leja M, Wex T, Malfertheiner P. Markers for gastric cancer premalignant lesions: where do we go? *Dig Dis* 2012;30(3):268-276. doi: 10.1159/000336990
66. Mandal S, Abebe F, Chaudhary J. -174G/C polymorphism in the interleukin-6 promoter is differently associated with prostate cancer incidence depending on race. *Genet Mol Res* 2014;13(1):139-151. doi: 10.4238/2014.january.10.5
67. Liu RY, Song X, Chen P, et al. Association between IL6 -174G/C and cancer: A meta-analysis of 105,482 individuals. *Exp Ther Med* 2012;3(4):655-664. doi: 10.3892/etm.2012.454
68. Zhang L, Du C, Guo X, et al. Interleukin-8-251A/T polymorphism and Helicobacter pylori infection influence risk for the development of gastric cardiac adenocarcinoma in a high-incidence area of China. *Mol Biol Rep* 2010;37(8):3983-3989. doi: 10.1007/s11033-010-0057-7
69. Li Z-W, Wu Y, Sun Y, et al. Inflammatory cytokine gene polymorphisms increase the risk of atrophic gastritis and intestinal metaplasia. *World J Gastroenterol* 2010;16(14):1788-1794. doi: 10.3748/wjg.v16.i14.1788
70. Yin YW, Hu AM, Sun QQ, et al. Association between interleukin-8 gene -251 T/A polymorphism and the risk of peptic ulcer disease: A meta-analysis. *Hum Immunol* 2013;74(1):125-130. doi: 10.1016/j.humimm.2012.09.006
71. Canedo P, Castanheira-Vale AJ, Lunet N, et al. The interleukin-8-251\*T/A polymorphism is not associated with risk for gastric carcinoma development in a Portuguese population. *Eur J Cancer Prev* 2008;17(1):28-32. doi: 10.3410/f.1102057.558030
72. Felipe AV, Silva TD, Pimenta CA, et al. Interleukin-8 gene polymorphism and susceptibility to gastric cancer in a Brazilian population. *Biol Res* 2012;45(4):369-374. doi: 10.4067/S0716-97602012000400007
73. Kim J, Cho YA, Choi IJ, et al. Effects of Interleukin-10 Polymorphisms, Helicobacter pylori Infection, and Smoking on the Risk of Noncardia Gastric Cancer. *PLoS ONE* 2012;7(1):1-8. doi: 10.1371/journal.pone.0029643
74. Akdogan RA, Ozgur O, Gucuyeter S, et al. A pilot study of Helicobacter pylori genotypes and cytokine gene polymorphisms in reflux oesophagitis and peptic ulcer disease. *Bratisl Lek Listy* 2014; 115(4): 221-228. doi: 10.4149/BLL\_2014\_046
75. Zeng X, Li Y, Liu T, et al. Diverse *H. pylori* strains, IL-10 promoter polymorphisms with high morbidity of gastric cancer in Hexi area of Gansu Province, China. *Mol Cell Biochem* 2012;362(1-2):241-248. doi: 10.1007/s11010-011-1149-y
76. Yuzhalin A. The role of interleukin DNA polymorphisms in gastric cancer. *Hum Immunol* 2011; 72(11): 1128-1136. doi: 10.1016/j.humimm.2011.08.003
77. Chen KF, Li B, Wei Yg, et al. Interleukin-10 -819 promoter polymorphism associated with gastric cancer among Asians. *J*

- Int Med Res 2010;38(1):1-8.
78. Brown LM. Helicobacter pylori: Epidemiology and Routes of Transmission. *Epidemiol Rev* 2000;22(2):283-287.
79. Malnick SD, Melzer E, Attali M, et al. Helicobacter pylori: Friend or foe? (27). *World J Gastroenterol* 2014;20:8979-8985. doi: 10.3748/wjg.v20.i27.8979
80. Yu M, Zhang XY, Yu Q. Detection of oral Helicobacter pylori infection using saliva test cassette. *Pak J Med Sci* 2015;31(5):1192-6. doi: 10.12669/pjms.315.7626
81. Cartágenes VD, Martins LC, Carneiro LM, et al. Helicobacter pylori in children and association with CagA strains in mother-child transmission in the Brazilian Amazon region. *Rev Soc Bras Med Trop* 2009;42(3):298-302. doi: 10.1590/S0037-86822009000300011
82. Rodrigues MN, Queiroz DMM, Braga ABC, et al. History of breastfeeding and Helicobacter pylori infection in children: results of a community-based study from northeastern Brazil. *Trans R Soc Trop Med Hyg* 2006;100(5):470-475. doi: 10.1016/j.trstmh.2005.08.001
83. Chen HL, Chen MJ, Shih SC, et al. Socioeconomic status, personal habits, and prevalence of Helicobacter pylori infection in the inhabitants of Lanyu. *J. of the Formosan Medical Association* 113(5):278-283, 2014. doi: 10.1016/j.jfma.2013.11.013
84. Sgouros SN, Bergele C. Clinical outcome of patients with Helicobacter pylori infection: the bug, the host, or the environment? *Postgrad Med J* 2006;82(967):338-342. doi: 10.1136/pgmj.2005.038273
85. Fialho AMN, Braga ABC, Braga Neto MB, et al. Younger Siblings Play a Major Role in Helicobacter pylori Transmission Among Children From a Low-Income Community in the Northeast of Brazil. *Helicobacter* 2010;15(6):491-496. doi: 10.1111/j.1523-5378.2010.00791
86. Dattoli VCC, Veiga RV, da Cunha SS, et al. Seroprevalence and Potential Risk Factors for Helicobacter pylori Infection in Brazilian Children. *Helicobacter* 2010;15(4):273-278. doi: 10.1111/j.1523-5378.2010.00766
87. Woodward M, Morrison C, Mccoll K. An investigation into factors associated with Helicobacter pylori infection. *J Clin Epidemiol* 2000;53(2):175-181. doi: 10.1016/S2222-1808(14)60326-1
88. Shikata K, Doi Y, Yonemoto K, et al. Population-based Prospective Study of the Combined Influence of Cigarette Smoking and Helicobacter pylori Infection on Gastric Cancer Incidence: the Hisayama Study. *Am J Epidemiol* 2008;168(12):1409-1415. doi: 10.1093/aje/kwn276